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# **Nanopolymer-Mediated Targeting Cancer with M1 Macrophage Polarizing Factors: A Unique Strategy to Fight against Cancer**

# **Ashok Chakraborty\*and Anil Diwan**

*AllExcel, Inc. Shelton, CT 06484, USA.*

**\* Correspondence:** Dr. Ashok Chakraborty, AllExcel, Inc. Shelton, CT 06484, USA.

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# **ABSTRACT**

*Macrophages are abundantly present, approx. 1010 cells, in almost all tissues of humans. They mainly function as phagocytic cells to make the human body disease-free from any eternal pathogens. The main two types of macrophages, M1 and M2 are known, of which M1 macrophages produces toxic NO, citrulline, tumor necrosis factor-α (TNF-α), and other pro-inflammatory cytokines, IL-12, IL- 23. M1 type while known as pro-inflammatory and also for their microbicidal and tumoricidal activities, the other type, M2 macrophages, are found to be antiinflammatory but tumor-promoting, one.*

*The unique features of M1 and M2 macrophages either as a tumoricidal or as a tumor promoting could be interested as a mechanism of tumor therapy, in future. Here, we review the strategies to exploit macrophages as therapeutic tools and targets in cancer therapy. In particular, the role of tumor-associated macrophages (TAMs) in cancer therapy will be focused.*

### **Keywords**

Macrophages, Cancer cells.

### **Abbreviations**

Arg-NPs: Arginine nanoparticles, β2m: β2-microglobulin, CTL: Cytotoxic T lymphocytes, MHC-II: Histocompatibility complex II class, INF: Interferon, IL-2: Interleukin-2, IL-12: Interleukin-12, JNK: C-Jun N-terminal kinase, LILRB-1: Leukocyte immunoglobulin-like receptor-1, LPS: Lipopolysaccharides, MAPK-JNK: mitogen-activated protein kinase/c-Jun NH2 terminal kinase, MHC: Major histocompatibility complex, NO: Nitric oxide, RIP-1: Receptor-interacting protein-1, TRAF-2: TNF receptor-associated factor-2, ROS: Reactive oxygen species, SIRP-α: Signal-regulatory protein-α, Siglec-10: Sialic-acidbinding Ig-like lectin-10, TNF-α: Tumor necrosis factor-α, TAMs: Tumor-associated macrophages, TFNR-1: TNF-α tumor necrosis factor receptor-1, TRADD: TFNR-1 associated with DEATH domain protein, Th-1/-2: T helper-1 /-2, TAM: Tumor associated macrophages.

### **Introduction**

Macrophages are motile cells, can enter the site of injury with

high destructive potential. In addition to killing pathogens and any foreign cells, macrophages are capable in presenting the main histocompatibility complex II class (MHC II) antigen [1]. Current cancer treatment strategies are not only includes radiotherapy, chemotherapy, or surgical re-section as but also opened up the era of targeted as well as immunotherapy. Modulation of TAMs by polarizing to M1 and activating their signaling system has emerged as a promising and novel immunotherapy for cancer therapy.

Cell-to-cell contact is the manner in which activated macrophages attack cancer cells [2,3], at the time activated macrophages attack cancer cells, the concentrations of chitotriosidase and protease on cell surface will be high enough to induce cancer cell lysis. A number of serine proteases, such as elastase, collagenase and plasminogen activator, are synthesized and secreted by activated macrophages, similar to chitotriosidases [4-7].

### **How Do Macrophages Recognize and Kill Cancer Cells? (Figure 1)**

- In fact, more than 50% tumor associated cells are macrophages [8].
- Macrophages can sense the differences of tumor cell-

membrane composition than normal cell membranes. One of such tumor markers include an increased content of phosphatidylserine. Other may include the altered glycan structures (or glycosylation), like, carcino-embryonic antigen Tn antigen on the tumor cell surfaces, which could be recognized by lectin-like receptors on the cell membranes of macrophages [9].

- Currently, the molecular mechanisms of the antitumor activity of M1-type macrophages are not fully understood.
- It is known that macrophages can kill cancer cells by several mechanisms, such as:
	- 1. Indirect killing by recruitment of other immune cells that can lyse the cancer cells,
	- 2. Cytolysis of cancer cells through antibody (Ab) dependent cellular cytotoxicity, and
	- 3. M1 macrophages can kill target cells directly by producing nitrosative/oxidative stress (NO/ROS), which induces DNA damage, cytotoxicity, and apoptosis [10- 13].
- M1 macrophages are known to promote also indirect cytotoxicity by activating other immune cells, such as NK cells and T cells [14-16].
- The innate immune receptor Dectin-1 is expressed on dendritic cells and macrophages which is critical to NK-mediated killing of tumor cells that express N-glycan structures at high levels [17].



**Figure 1:** Mechanisms of macrophage-mediate cancer cell killing: 1. Direct killing; 2. Cytolysisi through antibody dependent cellular cytotoxicity; 3. Indirect killing.

### **How to Stimulate M1-type of Macrophages in Cancer Cells**

- In the T helper 1 (Th1) based response, macrophages undergo M1-type activation, synthesizing chitotriosidases, protease, NO,  $H_2O_2$ , and other chemicals that kill invaders, such as fungi, viruses, and bacteria.
- Th1 response also can destroy Cancer cells. In this case interferon (INF), interleukin12 (IL12), IL2, and tumor necrosis factor (TNF) play major roles [18,19].
	- $\Box$  TNF-α tumor necrosis factor receptor 1 (TFNR1) signaling pathway [20].
	- $\Box$  TNF- $\alpha$  binds to its receptor TFNR1 associated with

DEATH domain protein (TRADD), receptor-interacting protein 1 (RIP1), and TNF receptor-associated factor 2 (TRAF2) to form Complex II.

- Complex II induces ROS production and activation of caspase-3 and caspase-7 [21,22], and lyse the cancer cells.
- $\Box$  TNF- $\alpha$  also induces cancer cell apoptosis through the MAPK-JNK pathway [21].
- However, In the T helper 2 (Th2) response, IL4 and IL10 play major roles, and macrophages undergo M2 type activation [18,19]. The mechanisms that control the immune response are not yet known. However, dendritic cells [23], CD4<sup>+</sup> / CD25<sup>+</sup> regulatory T cells [24-26], and T helper17 cells [27] are involved.

Because only the activated M1 macrophages produce 50kDa chitotriosidases, we need to convert the Th2 response to the Th1 response in patients with cancer. To successfully induce the Th1 response, three points need to be followed.

- $\Box$  The amount of the stimulator used to stimulate the Th1 response must be appropriately low, otherwise resistant may develop [28,29].
- $\Box$  Either below or higher of that special range, naive  $\Box$  cells would develop the Th2 response and produce a large amount of IL4 *and little IFN-y* [30].

### **Polarization of Tams and their Role in the TME**

- Theoretically, TAM could be differentiated into M1 phenotype macrophages by Th1 (IFN-γ, TNFα, and LPS et al.), and into M2 by Th2 (IL-4, IL-10, TGFβ1, and PGE2 et al.) cytokines [31] (Figure 2).
- M1 macrophages are characterized by the expression of HLA-DR and CD197, whereas M2 typically express scavenger receptor (CD163), CD209, mannose receptors (CD206), and CCL2, VEGF, and IL-10, etc. [32,33].
- A few sunsets of TAMs in the TME express CD86 and CD80 markers and are termed as M1-like TAMs, and typically exhibit anti-tumor effects [34].



**Figure 2:** Polarization of TAMS and their role in the TME.

### **Cancer Treatment Strategy**: **Targeting Tam Repolarization to M1 Type**

Targeting TAMs for cancer treatment has two main directions:

- $\Box$  To prevent macrophage recruitment to the TME; and
- $\Box$  To regulate TAM repolarization [35].
- $\Box$  Restriction of the infiltration of TAMs by blocking the CSF-1/ CSF-1 receptor and CCL2/CCR2 pathways showed some success, also.

### **A: STAT Signaling Pathways Drives M1 Polarization and cancer inhibition:**

- The mechanisms of TAM polarization are found to be correlated with several signaling pathways that includes C-Jun N-terminal kinase (JNK), PI3K/ Akt, JAK/ STAT and Notch signaling pathways [36].
- The JAK/STAT1/STAT6 signaling pathway can be activated by IFN-g [37], which can induce NAMPT-driven glycolysis, M1 polarization.
	- □ Inhibition of TAM-derived IL-6 can modulate CCL2 secretion, thus inhibits CD163<sup>+</sup> TAM polarization in colorectal cancer and attenuates tumor occurrence [38].

### **B: Inhibition of PI3Kg /AKT Signaling Pathway drives M1 Polarization and cancer inhibition**

- PI3K inhibitor, LY294002 effectively suppresses the polarization of M2 macrophages, and thereby their tumorigenic process [39].
- $\Box$  Further, the PI3Kg pathway-blocking drug BEZ successfully switched TAMs from M2 to M1 phenotype and inhibits pancreatic cancer progression [40].

## **C: Inhibition of CD47/SIRPa Signaling Pathway can stimulate M1 Polarization**

- □ The CD47/SIRPa *"do not eat me"* signal is of great interest in terms of the anti-phagocytic ability of macrophages.
- □ Anti-CD47 treatment has been reported to regulate the transformation of M2-like TAMs into the M1 phenotype *in vivo* [41].

Targeting the CD47/SIRPa axis results better prognosis in various cancers such as ovarian, breast and colorectal cancer [42] (Figure 3).



SIRP  $\alpha$ : Signal regulatory protein- $\alpha$ 

LILRB1 Gene - Leukocyte Immunoglobulin Like Receptor B1 PD1: Programmed cell death protein Siglec-10: Sialic acid binding Ig-like lectin 10

**Figure 3:** Anti-phagocytic checkpoints in the tumor microenvironment. The expression of "don't-eat-me" signals on tumor cells, including, CD47, PD-L1, MHC-1, and CD24, protect tumor cells from phagocytic clearance by interacting with their cognate receptors on macrophages.

# **D: Other Factors Involved in Macrophage Polarization**

- Lactic Acid and Tumor Acidosis Promotes M2 Macrophage Polarization and thus tumor invasion [43,44].
	- $\Box$  Whereas, M1 pro-inflammatory macrophages rely mainly on glycolysis, which in turn increases lactic acid levels [43].
	- $\Box$  Therefore, it is crucial to regulate the balance between lactate levels and the degree of tumor acidosis in the TME to inhibit immunosuppression.

# **Iron Promotes M1 Macrophage Polarization**

- $\Box$  Iron facilitates M1 polarization, while inhibits M2 activation, and exerting tumor immunotherapy effects.
- $\Box$  Further, Fe level can up-regulate the expression of CCL2, IL-1b, TNF-a, and IL-6, and thereby activates the NF-kB signaling pathway, and finally the M1/M2 macrophage polarization [45].

# **Phytochemicals and Macrophages Polarization**

 $\Box$  Curcumin blocks M2 polarization in microglial cells of the mouse brain and increase the M1 polarization by inducing STAT1 and NF-kB signaling pathways [46].

# **E: Targeting Macrophage-Derived and Cancer Cell-Derived Exosomes**

- Exosomes are typically between 30 and 150 nm in diameter in size, enclosed by a lipid membrane, which have been found to participate in the interaction between tumor cells and macrophages.
- Both, macrophage-derived and cancer cell-derived exosomes are associated with various signaling pathways that control the tumor immune evasion, metastasis, and drug resistance [47].
- For example, exosomal miR-138-5p derived from breast cancer cells suppresses the expression of KDM6B in macrophages and inhibits M1- related gene expression and thereby promoting breast cancer metastasis [48].
- Therefore, corrections of the ability of exosomes to target tumors and to overcome the biological barriers, like blood– brain barrier and gastrointestinal tract, are emerging as potential therapeutic strategies [47].

# **F: M2 macrophages are capable of active proliferation** *in situ*

- They express higher levels of several marker receptors including:
	- $\Box$  CD36 scavenger receptor for apoptotic cells,
	- □ CD206 mannose receptor,
	- $\Box$  CD301 receptor for galactose, and N-acetylglucosamine,
	- $\Box$  CD163 receptor for the hemoglobin–haptoglobin complex.
- Macrophages of this type are characterized by a low IL-12/ IL-10 secretion ratio.
- Furthermore, M2 phenotype macrophages are subdivided into M2a, M2b, M2c, and M2d subtypes depending on their markers and functions [49-53].
- The possibility of transformation of the M1 phenotype into M2 was shown with a range of cytokines and as a result of efferocytosis.
- The reverse transformation of the M2 phenotype into M1 is assumed with the development of obesity [54-56].

### **How Do Tumor Cells Bypass the Antitumor Effect of Macrophages?**

- Although macrophages exhibit tumoricidal and phagocytic activity of tumor cells *in vitro,* in reality, tumor cells can bypass the immune system and avoid the action of macrophages.
- CD47, PD-L1, major histocompatibility complex (MHC) class I, and CD24 are the molecules that are expressed by tumor cells and can inhibit the phagocytic activity of macrophages.
- In fact, signal-regulatory protein-α (SIRPα), PD-1, leukocyte immunoglobulin-like receptor 1 (LILRB1), and sialic-acidbinding Ig-like lectin-10 (Siglec-10), the respective binding receptors-45 are expressed on macrophage surface (Figure 3).
- **CD47:** The cancer cells often over-express the membranebound protein CD47, which is often called the *"don't-eatme"* signal. This signal suppresses the phagocytic activity of macrophages upon binding to SIRPα (signal regulatory protein  $α$ )-receptor present on phagocytes [57-59].
- Blocking CD47-SIRPα binding promotes phagocytosis of tumor cells by macrophages and induces antitumor responses in different xenograft models [57,60].
- **MHC1:** In addition, a second mechanism was found to play by the MHC class I component β2-microglobulin (β2m) that are expressed by cancer cells which can protect them from phagocytosis. Disruption of either MHC class I potentiates phagocytosis of tumor cells both *in vitro* and *in vivo* [61].
- **PD-1:** PD-1 is a membrane protein of the immunoglobulin superfamily that is involved in the cellular differentiation of immune cells. PD-1 and its ligands, PD-L1 and PD-L2, prevent the activation of T lymphocytes [10-13].

More recently, PD-1 has been found in TAMs, and its expression correlates with tumor growth [62]. PD-1- macrophages exhibit a higher level of phagocytic activity compared to PD-1<sup>+</sup> macrophages. Further, blockade of PD-1 or PD-L1 with Abs leads to increased phagocytosis of tumor cells and suppresses tumor growth. Thus, the PD-1 not only serves as a checkpoint for T cells but also contributes to the evasion of tumor cells from the killing by macrophages.

- **CD24:** Some tumor cells overexpress a glycosylated surface protein, CD24, which interacts with Siglec-10 on immune cells, and suppress inflammatory responses caused by tissue damage and avoid phagocytosis by macrophages expressing Siglec-10 [63-65].
- **Siglec-10:** Tumor-associated M2 macrophages express higher levels of Siglec-10 and are less phagocytic than M1 macrophages.

## **Use of Biologically Active Compounds for Macrophage Activation, Polarization, and Reprogramming**

**LPS:** It is well known that bacterial LPS can activate macrophages via Toll- like receptors [66]. However, the use

of LPS for the activation and polarization of macrophages for anticancer immunotherapy in patients is practically underscored due to the detrimental effect of LPS on the immune system. In this regard, the search for new methods to obtain stimulated macrophages continues.

For instance, human fibronectin and C-reactive protein have been studied as stimulants of macrophages *in vitro*  [67,68] as well as beta-1,3-D- glucan from yeast cell walls [69], lipophilic muramyldipeptide analogs [70], lipoprotein containing lipophilic muramyl tripeptide [71], lipopeptide analog of a fragment from the cell wall of gram-negative bacteria [72].

- Plant Extracts: An extract from Crasso-cephalum crepidioides, a plant distributed in Okinawa Islands (Japan), stimulated macrophages to enhance the expression of iNOs and increase the synthesis of NO, and activated antitumor activity against murine Sarcoma 180 (S-180) cancer cells [73].
- **Baicalein** (5,6,7-trihydroxyflavone), isolated from the Chinese herb Scutellaria baicalensis root, can block TGF-β1 via inhibiting PI3K/Akt pathway in M2 macrophages and repolarize them to a M1 phenotype in breast cancer tissues [74].
- An extract from the root of *Panax notoginseng* can re-educate M2-like macrophages toward M1 differentiation [75].
- **Ginsenoside Rb3** from *Panax ginseng* has protective functions against acute lung injury via M1/M2 phenotypic switch [76].
- **Emodin** (1,3,8-trihydroxy-6-methylanthraquinone) bidirectionally regulates both M1 and M2 phenotype programs via different mechanisms including suppression of STAT6 and C/EBPβ signaling to increase H3K27m3 on the M2 related genes. Emodin also restrains excessive M1 or M2 macrophages [77].
- **Osthole** [7-methoxy-8-(3-methyl- 2-butenyl)-2H-1benzopyran-2-one] is a coumarin member isolated from Cnidiummonnieri (Fructus Cnidii), found to decrease M2-like macrophages in pancreatic tumors by inhibition of STAT6 and p-ERK1/2-C/ EBP β [78].
- **Several "marine" compounds** can regulate macrophages and lead to the polarization of macrophages in both the M1 and M2 phenotypes [79]. Recent studies on the membranetype di-terpenoids isolated from soft coral species Briareum violaceum in Taiwan demonstrated suppressive effects on iNOS release from the cells, suggesting a potential to shift the M1 phenotype toward the M2 type [80].
- **Polysaccharides** isolated from gorgonian Pseudopterogorgia americana induces the expression of TNF-α, IL-6, and COX-2 in mouse macrophages, but had no effect on the expression of iNOS and NO production.

These compounds decrease expression of proinflammatory cytokines in LPS-activated macrophages, indicating a potential in reprogramming of macrophage polarization toward the M1-type [81].

The purine alkaloid homarine, a major metabolite found in water extracts of Portugal sea anemones, Actinia equine and *Anemonia sulcata*, has a great potential in modulating macrophage polarization [82].

- **Crustaceans of the order Decapoda** (crab, shrimp, prawn, and lobster) are a valuable source of plain polysaccharide and chitosan, which may promote the drug delivery targeting M1 or M2 macrophages [83].
- **Triterpene glycosides** are capable of activating macrophages both *in vivo* and *in vitro* and polarizing them into the M1 phenotype by affecting the purinergic P2X4 receptors pathway [84-86]. Increased ROS and NO levels are found in cells treated with triterpene glycosides.

### **Car-Engineered Macrophages Act Like M1 Cells (Car-M Cells)**

• One of the promising approaches in cell therapy for cancer is the use of CAR initially applied to modify T cells. The CAR-engineered macrophages express pro-inflammatory cytokines and chemokines, convert M2 macrophages to M1, activate the antigen presentation mechanism, recruit and present antigen to T cells, and resist immunosuppressive cytokines. In addition, they exhibit pronounced antigenspecific phagocytosis and *in vitro* tumor suppression.

In two mouse models of solid tumor xenograft, a single infusion of CAR-macrophages reduced the growth of human tumors and increased the overall survival of tumor-bearing animals. In humanized mouse models, it was also shown that genetically modified macrophages induce a pro- inflammatory tumor microenvironment and enhance the antitumor activity of T cells [87]. In a series of recent studies, it was found that the suppression of SIRPα on macrophages from the bone marrow of mice and humans leads to the blocking of recognition of its own "marker of self," CD47, on all other cells.

**• An integrated nanotechnology/biology strategy for cancer immunotherapy** that uses arginine nanoparticles (Arg-NPs) to deliver CRISPR-Cas9 gene editing machinery into cells to generate SIRP-α knockout macrophages and block its binding to CD47 was recently reported [88]. The NP system efficiently co-delivers single guide RNA and Cas9 protein required to knockout the **"don't-eat-me"** signal in RAW 264.7 macrophages. Turning off this signal increased the phagocytosis of human osteosarcoma U2OS cancer cells by fourfold.

### **Use of Nanoparticles (NPS) As a Carrier for M1 Polarizing Factors**

NPs are very tiny particles (<100 nanometers) made up of latex, polymers, ceramic materials, metals, and carbon particles. Their surfaces could be hydrophilic or hydrophobic, and exhibit surface charges and specific ligands, reference for the treatment of various clinical diseases [89]. Other synthetic NPs could have been made using liposomes [90,91], polylactic-co-glycolic acid (PLGA) [92,93], chitosan [94], silica [95], and dextran materials [96]. Applications of NPs in Medical science are increasing due to their physicochemical properties, e.g., size, shape, structure, chemical composition, morphology, and surface properties, etc. Now-a-days, many researchers are using formulations with small

molecules and NPs, such as Toll-like receptor (TLR) agonists, cytokines, antibodies, and RNAs, for macrophage repolarization [97]. Polymeric NPs synthesized with an IL-12 cytokine promote the conversion of M2 to M1-type, and thus could be used for cancer immunotherapy [98]. In 2018, Rodell et al. showed that R848, an agonist of TLR7 and TLR8, shift TAM to an M1 phenotype, and controls the tumor growth [99,100]. Furthermore, since immune cells express mannose receptors, mannose carbohydrate can also be employed to target macrophages [101,102]. Zhao et al. synthesized the albumin NPs having dual ligands, one, a transferrin receptor (TfR)-binding peptide T12, and the second is the mannose. They showed that this therapeutic efficiently can polarize pro-tumor M2 to anti-tumor M1 and inhibit the glioma cell proliferation successfully [103]. Most of mannose are now being applied for macrophage re-polarization as a ligand to target macrophages.

Interestingly, inhibition of CD40 also leads to IL-12 upregulation, which can repolarize TAMs into M1 macrophages. Similarly, inhibition of NF-κB signaling pathway can switch TAMs into M1 macrophages and block tumor cell's growth. Therefore, these approaches have a potential importance in cancer therapy [104]. Another strategy is to make a pH-sensitive NPs. Tumor pH ranges generally from 6.5 to 6.8 while the pH in healthy tissues is 7.4. NPs, like poly(acryl amide) (PAAm), micelles and liposomes can release drugs through protonation or deprotonation designed to be pH-sensitive and bypass the normal tissues [89,105-109].

**Theracure Biopolymer Nanovehicles as an Antcancer Regimen** TheraCour platform polymer is a self-assembling, uniform, tailorable linear homopolymer that comprises polyethylene glycol (PEG) within its monomer unit which is heterochemically functionalized with a specially designed linker unit so that covalently connected aliphatic chains are suspended from it and separately site-targeting ligands are also covalently attached to it [110-112] (Figure 4).



**Figure 4:** Schematic Presentation of Nanovehicles.

This simple scheme results in a polymer that is like a half-biological membrane, self-assembles into micelles with hydrophobic, flexible core region made of the lipid chains. The hydrophilic ligands directing outwards into the aqueous milieu are ready to seek their partners, connected together by the corona of PEG. Upon binding of the TheraCour polymeric micelle to a cellular receptor may initiate lipid mixing of the flexible pendant interior lipid chains of the micelle with the flexible lipids of the cell membrane, leading to passive fusion. Alternatively, receptor-mediated endocytosis can take place at properly chosen receptors. These processes would result in site-specified or address- targeted delivery of the encapsulated drug payload content of the micelle. The graphical model of anticancer mechansim of TheraCour platform technology are shown in Figure 4. [For Review, 113].

This polymer can effectively encapsulate many types of chemotherapeutic APIs, target the cancer cell based on the selected ligand, and thereby result in effective anticancer activity. Recently it was shown that the cell proliferation of two lung cancer cell lines (A549 and H441), and two breast cancer cell lines (SKBR3 and BT474) are mostly inhibited by a folate-targeted TheraCour polymer delivering API (camptothecin, CPT) [113].

### **Discussion**

Macrophages are the "big eaters" in the animals and humans body capable of engulfing any foreign bodies including bacteria, dead cells, and other particles that are toxic to the body. They also activate T- and B-lymphocytes by antigens presentation.

Macrophages are present in virtually every organ/tissue where they act as the first line of immune defense against pathogens and play an important role in maintaining tissue homeostasis. Classically activated macrophages (M1 phenotype) are stimulated by lipopolysaccharides (LPS) as well as IFN-γ in conjunction with LPS. The "activated macrophage" has an increased ability to kill microorganisms or tumor cells [114].

Polarization toward the M1 direction is accompanied by the secretion of pro- inflammatory mediators. They express receptors for IL-1 (IL-1R1), TLR, and co- stimulatory molecules, the activation of which ensures the amplification of the inflammatory response [115-118]. IL-12 secreted by M1 macrophages also plays a key role in Th1 polarization, whereas IL-1β and IL-23 direct the immune response along the Th17 pathway [115-118].

Alternative activation of macrophages (M2 macrophages) is observed when the cells are stimulated by interleukins, glucocorticoids, immune complexes, TLR agonists, etc. Briefly, M2 phenotype macrophages preferentially activate the motility of cancer cells [119], promote metastasis in stromal and perivascular areas [120], and stimulate angiogenesis in a vascular and perinecrotic hypoxic areas [121].

In theory, accumulation of macrophages in tumor activate the stroma [122] and ECM remodeling, including lysyl oxidase, MMP9, type IV collagen activation [123]. To make sense of macrophages polarization in cancer remission, studies could be

conducted to estimate the number of macrophages sub-type in an outside and inside of the tumor during their different stages of treatment.

Therefore, a complete understanding of nanomaterials interaction with distinct polarized macrophage phenotypes, is important to translate the nanomedicines for clinical purposes. The major points to be considered are:

- Classically activated M1 macrophages are microbicidal and pronflammatory while alternatively activated M2 macrophages are predominantly immune modulators and antiinflammatory [124].
- The differential uptake methods of nanoparticles by M1 and M2-type macrophages are complex, and involves cytoskeletal remodeling, membrane fusion and vesicular transport [125- 127].

Recently it was reported that the polarization of macrophages towards the M1 phenotype resulted an increased uptake ability of non-PEGylated nanoparticles compared to their M2 counterparts [125]. In contrast, inhibition of CD47-SIRPα by anti-CD47 antibodies produced a higher pro-phagocytosis of cancer cells by M1-type as compared to M2-type macrophages [128]. However, how the M1/M2 polarization system works in the tissue-specific macrophages such as microglia in the central nervous system to cause neuro-inflammation remains unknown [129].

In brief, activated macrophage populations possess unique pro- and anti- inflammatory type, which play an important role in immune regulation as well as in disease pathology. During strokes, for example, M1 macrophages becomes active and promote inflammation after ischemic injury, release cytotoxic cytokines and ultimately neuronal death [130]. In contrast, M2 like tumor-associated macrophages promote immune suppression and facilitate tumor invasion [2]. Therefore, the understanding of the mechanism(s) how the nano-materials interact with specific macrophage phenotypes and the ability to design nano-materials for selective targeting to those macrophage subpopulations are crucial parts in designing the nano-medicines. Combining all the above information a conclusive picture therefore can be drawn to combat cancer as follows (Figure 5).



**Figure 5:** NPs mediated Macrophage Polarization and their Effects.

### **Conclusion**

- In summary, the two functionally distinct macrophage subtypes, M1 and M2, are the key modulators in host immune system against tumors.
- Based on the appropriate system they could act like a "friend or foe," of the tumor cells either to assist or destroy their growth by various mechanisms.
- In direct contact with tumor cells, macrophages attack them with cytotoxic molecules followed by phagocytizing the remnants of the tumor cells.
- In addition, they can activate the production of cytotoxic molecules by cytotoxic T lymphocytes (CTL) and NK cells followed by the production of their specific antibodies by B-lymphocytes.
- However, tumor can "fool" macrophages, and bypass the M1 effect and recruit M2 type for tumor growth.
- In this regard, scientists are trying to find new ways of additional activation of a larger number of macrophages and programming to their M1 phenotype.

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#### **References**

- 1. Plowden J, Renshaw Hoelscher M, Engleman C, et al. Innate immunity in aging: impact on macrophage function. Aging Cell. 2004; 3: 161-167.
- 2. Gordon S, Newman W, Bloom B. Macrophage proteases and rheumatic diseases: regulation of plasminogen activator by thymus-derived lymphocytes. Agents Actions. 1978; 8: 19-26.
- 3. Somers SD, Whisnant CC, Adams DO. Quantification of the strength of cell adhesion: the capture of tumor cells by activated murine macrophages proceeds through two distinct stages. J Immunol. 1986; 136: 1490-1496.
- 4. Harwix S, Andreesen R, Ferber E, et al. Human macrophages secrete a tumoricidal activity distinct from tumor necrosis necrosis factor and reactive nitrogen intermediates. Res Immunol. 1992; 143: 89-94.
- 5. Adams DO, Kao KJ, Farb R, et al. Effector mechanism of cytolytically activated macrophages II. Secretion of a cytolytic factor by activated macrophages and its relationship to secreted neutral proteases. J Immunol. 1980; 124: 293-300.
- 6. Sundsmo JS, Chin JR, Papin RA, et al. Factor B, the complement alternative pathway serine proteinase is a major constitutive protein synthesized and secreted by resident and elicited mouse macrophages. J Exp Med. 1985; 161: 306-322.
- 7. Adams DO. Effector mechanisms of cytolytically activated macrophages. I. Secretion of neutral proteases and effect of protease inhibitors. J Immunol. 1980; 124: 286-292.
- 8. Dolgin E. Cancer-eating immune cells kitted out with CARs. Nat Bio- technol. 2020; 38: 509-511.
- 9. Klimp AH, de Vries EG, Scherphof GL, et al. A potential role of macrophage activation in the treatment of cancer. Crit Rev Oncol Hematol. 2002; 44: 143-161.
- 10. Garban HJ, Bonavida B. Nitric oxide sensitizes ovarian tumor cells to Fasinduced apoptosis. Gynecol Oncol. 1999; 73: 257- 264.
- 11. Lee SY, Rim Y, McPherson DD, et al. A novel liposomal nanomedicine for nitric oxide delivery and breast cancer treatment. Biomed Mater Eng. 2014; 24: 61-67.
- 12. Mojic M, Mijatovic S, Maksimovic Ivanic D, et al. Therapeutic potential of nitric oxide- modified drugs in colon cancer cells. Mol Pharmacol. 2012; 82:700-710.
- 13. Rahat MA, Hemmerlein B. Macrophage-tumor cell interactions regulate the function of nitric oxide. Front Physiol. 2013; 4: 144.
- 14. Darcy PK, Neeson P, Yong CS, et al. Manipulating immune cells for adoptive immunotherapy of cancer. Curr. Opin Immunol. 2014; 27: 46-52.
- 15. Fridlender ZG, Albelda SM. Modifying tumor-associated macro- phages: an important adjunct to immunotherapy. Oncoimmunology. 2013; 2: 26620.
- 16. Seledtsov VI, Goncharov AG, Seledtsova GV. Clinically feasible approaches to potentiating cancer cell-based immunotherapies. Hum Vaccin Immunother. 2015; 11: 851-869.
- 17. Chiba S, Ikushima H, Ueki H, et al. Recognition of tumor cells by Dectin-1 orchestrates innate immune cells for antitumor responses. Elife. 2014; 3: 04177.
- 18. Sica A, Larghi P, Mancino A, et al. Macrophage polarization in tumor progression. Semin Cancer Biol. 2008; 18: 349-355.
- 19. Onoe K, Yanagawa Y, Minami K, et al. Th1 or Th2 balance regulated by interaction between dendritic cells and NKT cells. Immunol Res. 2007; 38: 319-332.
- 20. Hsu H, Shu HB, Pan MG, et al. TRADD–TRAF2 and TRADD– FADD interactions define two distinct TNF receptor 1 signal transduction pathways. Cell. 1996; 84: 299-308.
- 21. Kim JJ, Lee SB, Park JK, et al. TNF-alpha-induced ROS production triggering apoptosis is directly linked to Romo1 and Bcl-X(L). Cell Death Differ. 2010; 17: 1420-1434.
- 22. Wang X, Lin Y. Tumor necrosis factor and cancer, buddies or foes?. Acta Pharmacol Sin. 2008; 29: 1275-1288.
- 23. Yamazaki S, Steinman RM. Dendritic cells as controllers of antigen-specific 22 Foxp3+ regulatory T cells. J Dermatol Sci. 2009; 54: 69-75.
- 24. Zhen Y, Zheng J, Zhao Y. Regulatory CD4+CD25+ T cells and macrophages: communication between two regulators of effector T cells. Inflamm Res. 2008; 57: 564-570.
- 25. Liu G, Ma H, Qiu L, et al. Phenotypic and functional switch of macrophages induced by regulatory CD4+/ CD25+ T cells in mice. Immunol Cell Biol. 2011; 89: 130-142.
- 26. Tiemessen MM, Jagger AL, Evans HG, et al. CD4+/CD25+/ Foxp3+ regulatory T cells induce alternative activation of human monocytes/macrophages. PNAS USA. 2007; 104: 19446-19451.
- 27. Xu S, Cao X. Interleukin17 and its expanding biological functions. Cell Mol Immunol. 2010; 7: 164-174.
- 28. Bretscher PA, Wei G, Menon JN, et al. Establishment of stable, cell-mediated immunity that makes susceptible mice resistant to Leishmenania major. Science. 1992; 257: 539-542.
- 29. Bretscher PA, Ogunremi O, Menon JN. Distinct immunological states in murine cutaneous leishmaniasis by immunizing with different amounts of antigen: the generation of beneficial, potentially harmful, harmful and potentially extremely harmful states. Behring Inst Mitt. 1997; 98: 153-159.
- 30. Lentinan AT, Fenishel RL, Chirgis MA. Immune Modulation Agents and Their Mechanism. New York: Marcel Dekker. 1984; 63-77.
- 31. Enderlin Vaz da Silva Z, Lehr HA, Velin D. *In vitro* and i*n vivo* repair activities of undifferentiated and classically and alternatively activated macrophages. Pathobiology. 2014; 81: 86-93.
- 32. Gao L, Wang FQ, Li HM, et al. CCL2/EGF positive feedback loop between cancer cells and macrophages promotes cell migration and invasion in head and neck squamous cell carcinoma. Oncotarget. 2016; 7: 87037-87051.
- 33. Feriotti C, Loures FV, Frank de Araujo E, et al. Mannosylrecognizing receptors induce an M1-like phenotype in macrophages of susceptible mice but an M2- like phenotype in mice resistant to a fungal infection. PLoS One. 2013; 8: 54845.
- 34. Shu Y, Cheng P. Targeting Tumor-Associated Macrophages for Cancer Immunotherapy. Biochim Biophys Acta Rev Cancer. 2020; 1874: 188434.
- 35. Liu KX, Joshi S. "Re-Educating" Tumor Associated Macrophages as a Novel Immunotherapy Strategy for Neuroblastoma. Front Immunol. 2020; 11: 1947.
- 36. Zhou D, Huang C, Lin Z, et al. Macrophage polarization and function with emphasis on the evolving roles of coordinated regulation of cellular signaling pathways. Cell Signal. 2014; 26: 192-197.
- 37. Huffaker TB, Ekiz HA, Barba C, et al. A Stat1 Bound Enhancer Promotes Nampt Expression and Function Within Tumor Associated Macrophages. Nat Commun. 2021; 12: 2620.
- 38. Hou, S, Zhao Y, Chen J, et al. Tumor-associated macrophages in colorectal cancer metastasis: molecular insights and translational perspectives. J Transl Med. 2024; 22: 1-14.
- 39. Wang Y, Lyu Z, Qin Y, et al. FOXO1 Promotes Tumor Progression by Increased M2 Macrophage Infiltration in Esophageal Squamous Cell Carcinoma. Theranostics. 2020; 10: 11535-11548.
- 40. Li M, Li M, Yang Y, et al. Remodeling Tumor Immune Microenvironment via Targeted Blockade of PI3K-g and CSF-1/CSF-1R Pathways in Tumor Associated Macrophages for Pancreatic Cancer Therapy. J Control Release. 2020; 321: 23-35.
- 41. Zhang M, Hutter G, Kahn SA, et al. AntiCD47 Treatment Stimulates Phagocytosis of Glioblastoma by M1 and M2 Polarized Macrophages and Promotes M1 Polarized Macrophages i*n Vivo*. PloS One. 2016; 11: 0153550.
- 42. Yang H, Shao R, Huang H, et al. Engineering Macrophages to

Phagocytose Cancer Cells by Blocking the CD47/SIRPa Axis. Cancer Med. 2019; 8: 4245-4253.

- 43. Van den Bossche J, O Neill LA, Menon D. Macrophage Immunometabolism: Where Are We (Going)?. Trends Immunol. 2017; 38: 395-406.
- 44. Bohn T, Rapp S, Luther N, et al. Tumor Immunoevasion via Acidosis-Dependent Induction of Regulatory TumorAssociated Macrophages. Nat Immunol. 2018; 19: 1319-1329.
- 45. Handa P, Thomas S, Morgan-Stevenson V, et al. Iron Alters Macrophage Polarization Status and Leads to Steatohepatitis and Fibrogenesis. J Leukoc Biol. 2019; 105: 1015-1026.
- 46. Koh YC, Yang G, Lai CS, et al. Chemopreventive Effects of Phytochemicals and Medicines on M1/M2 Polarized Macrophage Role in Inflammation-Related Diseases. Int J Mol Sci. 2018; 19: 2208.
- 47. Shao J, Zaro J, Shen Y. Advances in Exosome-Based Drug Delivery and Tumor Targeting: From Tissue Distribution to Intracellular Fate. Int J Nanomed. 2020; 15: 9355-9371.
- 48. Xun J, Du L, Gao R, et al. Cancer-Derived Exosomal miR-138-5p Modulates Polarization of Tumor-Associated Macrophages Through Inhibition of KDM6B. Theranostics. 2021; 11: 6847-6859.
- 49. Gratchev A, Kzhyshkowska J, Utikal J, et al. Interleukin-4 and dexamethasone counter regulate extracellular matrix remodelling and phagocytosis in type-2 macrophages. Scand J Immunol. 2005; 61: 10-17.
- 50. Kreider T, Anthony RM, Urban JF, et al. Alternatively activated macrophages in helminth infections. Curr Opin Immunol. 2007; 19: 448-453.
- 51. Trial J, Cieslik KA, Haudek SB, et al. Th1/M1 conversion to Th2/M2 responses in models of inflammation lacking cell death stimulates maturation of monocyte precursors to fibroblasts. Front Immunol. 2013; 4: 287.
- 52. Xiong W, Frasch SC, Thomas SM, et al. Induction of TGF-β1 synthesis by macrophages in response to apoptotic cells requires activation of scavenger receptor CD36. PLoS One. 2013; 8: 72772.
- 53. Cheng H, Wang Z, Fu L, et al. Macrophage polarization in the development and progression of ovarian cancers: an overview. Front Oncol. 2019; 9: 421.
- 54. Gong D, Shi W, Yi S, et al. TGF- βsignaling plays a critical role in promoting alternative macrophage activation. BMC Immunol. 2012; 13: 31.
- 55. Xiong W, Frasch SC, Thomas SM, et al. Induction of TGF-β1 synthesis by macrophages in response to apoptotic cells requires 27 activation of scavenger receptor CD36. PLoS One. 2013; 8: 72772.
- 56. Ylöstalo JH, Bartosh TJ, Coble K, et al. Human mesenchymal stem/stromal cells (hMSCs) cultured as spheroids are self-activated to produce prostaglandin E2 (PGE2) that directs stimulated macro-phages into an anti-inflammatory phenotype. Stem Cells. 2012; 30: 2283-2296.
- 57. Majeti R, Chao MP, Alizadeh AA, et al. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. Cell. 2009; 138: 286-299.
- 58. Weiskopf K, Ring AM, Ho CC, et al. Engineered SIRPalpha variants as immunotherapeutic adjuvants to anticancer antibodies. Science. 2013; 341: 88-91.
- 59. Oldenborg PA, Gresham HD, Lindberg FP. CD47-signal regulatoryprotein alpha (SIRPalpha) regulates Fcgamma and complement receptormediated phagocytosis. J Exp Med. 2001; 193: 855-862.
- 60. Willingham SB, Volkmer JP, Gentles AJ, et al. The CD47 signal regulatory protein alpha (SIRPa) interaction is a therapeutic target for human solid tumors. Proc Natl Acad Sci USA. 2012; 109: 6662-6667.
- 61. Barkal AA, Weiskopf K, Kao KS, et al. Engagement of MHC class I by the inhibitory receptor LILRB1 suppresses macrophages and is a target of cancer immunotherapy. Nat Immunol. 2018; 19: 76-84.
- 62. Gordon SR, Maute RL, Dulken BW, et al. PD-1 expression by tumour-associated macrophages inhibits phagocytosis and tumor immunity. Nature. 2017; 545: 495-499.
- 63. Barkal AA, Brewer RE, Markovic M, et al. LCD24 signalling through macrophage Siglec-10 is a target for cancer immunotherapy. Nature. 2019; 572: 392-396.
- 64. Chen GY, Brown NK, Zheng P, et al. Siglec-G/10 in selfnonself discrimination of innate and adaptive immunity. Glycobiology. 2014; 24: 800-806.
- 65. Chen GY, Tang J, Zheng P, et al. CD24 and Siglec-10 selectively repress tissue damage-induced immune responses. Science. 2009; 323: 1722-1725.
- 66. Hancock RE, Nijnik A, Philpott DJ. Modulating immunity as a therapy for bacterial infections. Nat Rev Microbiol. 2012; 10: 243-254.
- 67. Perri RT, Kay NE, McCarthy J, et al. Fibronectin enhances *in vitro* monocyte–macrophage-mediated tumoricidal activity. Blood. 1982; 60: 430-435.
- 68. Zahedi K, Mortensen RF. Macrophage tumoricidal activity induced by human C-reactive protein. Cancer Res. 1986; 46: 5077-5083.
- 69. Bögwald J, Johnson E, Seljelid R. The cytotoxic effect of mouse macrophages stimulated in vitro by a beta-1,3-Dglucan from yeast cell walls. Scand J Immunol. 1982; 15: 297-304.
- 70. Phillips NC, Chedid L, Bernard JM, et al. Induction of murine macrophage tumoricidal activity and treatment of experimental pulmonary metastases by liposomes containing lipophilic muramyl-dipeptide analogs. J Biol Response Mod. 1987; 6: 678-691.
- 71. Shaw JM, Futch WS Jr, Schook LB. Induction of macrophage anti- tumor activity by acetylated low density lipoprotein containing lipophilic muramyl tripeptide. Proc Natl Acad Sci USA. 1988; 85: 6112-6116.
- 72. Utsugi T, Dinney CP, Killion JJ, et al. In situ activation of

mouse macrophages and therapy of spontaneous renal cell cancer metastasis by liposomes containing the lipopeptide CGP 31362. Cancer Immunol Immunother. 1991; 33: 375-381

- 73. Tomimori K, Nakama S, Kimura R, et al. Antitumor activity and macrophage nitric oxide producing action of medicinal herb. Crassocephalum crepidioides. BMC Complement Altern Med. 2012; 12: 78.
- 74. Zhao X, Qu J, Liu X, et al. Baicalein suppress EMT of breast cancer by mediating tumor-associated macrophages polarization. Am J Cancer Res. 2018; 8: 1528-1540.
- 75. Kim B, Kim EY, Lee EJ, et al. Panax notoginseng inhibits tumor growth through activating macrophage to M1 polarization. Am J Chin Med. 2018; 46: 1369-1385.
- 76. Yang J, Hang J, Li S, et al. Ginsenoside Rg3 30 attenuates lipopolysaccharide-induced acute lung injury via MerTKdependent activation of the PI3K/AKT/mTOR pathway. Front Pharmacol. 2018; 9: 850.
- 77. Wang B, Botao Hang J, Li S, et al. Osthole inhibits pancreatic cancer progression by directly exerting negative effects on cancer cells and attenuating tumor-infiltrating M2 macrophages. J Pharmacol Sci. 2018; 137: 290-298.
- 78. Wang JB, Zhao HP, Zhao YL, et al. Hepato- toxicity or hepatoprotection? Pattern recognition for the paradoxical effect of the Chinese herb Rheum palmatum L. in treating rat liver injury. PLoS One. 2011; 6: 24498.
- 79. Dolmatova LS, Dolmatov IY. Different macrophage type triggering as target of the action of biologically active substances from marine invertebrates. Mar Drugs. 2020; 18: 37.
- 80. Huang PC, Lin WS, Peng BR, Chang YC, Fang LS, Li GQ. New furanocembranoids from Briareumviolaceum. Mar Drugs. 2019; 17: 214.
- 81. Chernikov OV, Chiu HW, Li LH, et al. Polysacharides from Pseudopterogorgia americana modu- lates immune response in macrophages. Vestnik FEB RAS. 2018; 6: 103.
- 82. Costa Silva T, Branquinho de Andrade P, Paiva Martins F, et al. *In vitro* anti-inflammatory and cytotoxic effects of aqueous extracts from the edible sea anemones Anemonia sulcata and Actinia equina. Int J Mol Sci. 2017; 18: 653.
- 83. Komohara Y, Fujiwara Y, Ohnishi K, et al. Tumor-associated macrophages: potential therapeutic targets for anti-cancer therapy. Adv Drug Deliv Rev. 2016; 99: 180-185.
- 84. Aminin D. Immunomodulatory properties of sea cucumber triterpene glycosides. Springer; 2014; 3: 1-17.
- 85. Aminin D, Pislyagin E, Astashev M, et al. Glycosides from edible sea cucumbers stimulate macrophages via purinergic receptors. Sci Rep. 2016; 6: 39683.
- 86. Pislyagin EA, Manzhulo IV, Gorpenchenko TY, et al. Cucumarioside A2-2 causes macro- phage activation in mouse spleen. Mar Drugs. 2017; 15: 341.
- 87. Klichinsky M, Ruella M, Shestova O, et al. Human chimeric antigen receptor macrophages for cancer immuno-therapy. Nat Biotechnol. 2020; 38: 947-953.
- 88. Ray M, Lee YW, Hardie J, et al. CRISPRed-macrophages for cell-based cancer immunotherapy. Bioconjug Chem. 2018; 29: 445-450.
- 89. Sun T, Zhang YS, Pang B, et. al. Engineered nanoparticles for drug delivery in cancer therapy. Angew Chemie. 2014; 53: 12320-12364.
- 90. Nguyen TX, Huang L, Gauthier M, et al. Recent advances in liposome surface modification for oral drug delivery. Nanomedicine. 2016; 11: 1169-1185.
- 91. Ren H, He Y, Liang J, et al. Role of liposome size surface charge and PEGylation on rheumatoid arthritis targeting therapy. ACS Appl Mater Interfaces. 2019; 11: 20304-20315.
- 92. Danhier F, Ansorena E, Silva JM, et al. PLGA-based nanoparticles an overview of biomedical applications. J Controlled Release. 2012; 161: 505-522.
- 93. Acharya S, Sahoo SK. PLGA nanoparticles containing various anticancer agents and tumour delivery by EPR effect. Adv Drug Deliv Rev. 2011; 63: 170-183.
- 94. Rao W, Wang H, Han J, et al. Chitosan- decorated doxorubicinencapsulated nanoparticle targets and eliminates tumor reinitiating cancer stem-like cells. ACS Nano. 2015; 9: 5725-5740.
- 95. Diab R, Canilho N, Pavel IA, et al. Silica-based systems for oral delivery of drugs macromolecules and cells. Adv Colloid Interface Sci. 2017; 249: 346-362.
- 96. Ma L, Liu TW, Wallig MA, et al. Efficient targeting of adipose tissue macrophages in obesity with polysaccharide nanocarriers. ACS Nano. 2016; 10: 6952-6962.
- 97. Van Dalen FJ, Van Stevendaal M, Fennemann FL. Molecular repolarization of 33 tumor-associated macrophages. Molecules. 2018; 24: 9.
- 98. Wang Y, Lin YX, Qiao SL, et al. Polymeric nanoparticles promote macrophage reversal from M2 to M1 phenotypes in the tumor microenvironment. Biomaterials. 2017; 112: 153-163.
- 99. Rodell CB, Arlauckas SP, Cuccarese MF, et al. TLR7/8 agonist-loaded nanoparticles promote the polarization of tumor-associated macrophages to enhance cancer immunotherapy. Nat Biomed Eng. 2018; 2: 578-588.
- 100.Cully M. Cancer re-educating tumor-associated macrophages with nanoparticles. Nat Rev Drug Disc. 2018; 17: 468.
- 101.Irache JM, Salman HH, Gamazo C, et al. Mannose-targeted systems for the delivery of therapeutics. Expert Opin Drug Deliv. 2008; 5: 703-724.
- 102.Wang T, Zhang J, Hou T, et al. Selective targeting of tumor cells and tumor associated macrophages separately by twinlike core- shell nanoparticles for enhanced tumor-localized chemoimmunotherapy. Nanoscale. 2019; 11: 13934-13946.
- 103.Zhao P, Wang Y, Kang X, et al. Dual-targeting biomimetic delivery for antiglioma activity via remodeling the tumor microenvironment and directing macrophage-mediated immunotherapy. Chem Sci. 2018; 9: 2674-2689.
- 104.Fong CH, Bebien M, Didierlaurent A, et al. An antiinflammatory role for IKK-β through the inhibition of classical macrophage activation. J Exp Med. 2008; 205: 1269-1276.
- 105.Fukumura D, Jain RK. Tumor microenvironment abnormalities causes consequences and strategies to normalize. J Cell Biochem. 2007; 101: 937-949.
- 106.Kost J, Langer R. Responsive polymeric delivery systems. Adv Drug Deliv Rev. 2001; 46: 125-148.
- 107.Kim JO, Kabanov AV, Bronich TK. Polymer micelles with cross-linked polyanion core for delivery of a cationic drug doxorubicin. J Control Release. 2009; 138: 197-204.
- 108.Lo CL, Huang CK, Lin KM, et al. Mixed micelles formed from graft and diblock copolymers for application in intracellular drug delivery. Biomaterials. 2007; 28: 1225-1235.
- 109.Borchert U, Lipprandt U, Bilang M, et al. pH-induced release from P2VPPEO block copolymer vesicles. Langmuir. 2006; 22: 5843-5847.
- 110.https://www.bloomberg.com/press-releases/2021-03-09/ nanoviricides-incpan-coronavirus-covid-19-drug-candidatesare-highly-effective-in-pre-clinicalanimal-studies-i.
- 111.Barton RW, Tatake JG, Diwan AR. Nanoviricides- A Novel Approach to Antiviral Therapeutics. Bionanotechnology II. Taylor and Francis Group. 2011; 141-154.
- 112.Barton RW, Tatake JG, Diwan AR. Nanoviricides Targeted Anti-Viral Nanomaterials Handbook of Clinical Nanomedicine Nanoparticles Imaging Therapy and Clinical Applications. Jenny Stanford Publishing. 2016; 24: 1039-1046.
- 113.Anil Diwan, Jayant Tatake, Ashok Chakraborty. Therapeutic Uses of TheraCour™ Polymeric Nanomicelles Against Cancer Infectious Diseases and More. Book: Nanomaterials for Cancer Detection Using Imaging Techniques and Their Clinical Applications. Springer Nature. 2022.
- 114.Lavin Y, Mortha A, Rahman A, et al. Regulation of macrophage development and function in peripheral tissues. Nat Rev Immunol. 2015; 15: 731-744.
- 115.Mantovani A, Sica A, Sozzani S, et al. The chemokine system in diverse forms of macrophage activation and polarization. Trends Immunol. 2004; 25: 677-686.
- 116.Chen X, Wenke Z, Xu W, et al. Granulin exacerbates lupus nephritis via enchanсing macrophage M2b polarization. PLoS One. 2013; 8: 65542.
- 117.Graff JW, Dickson AM, Clay G, et al. Identifying functional microRNAs in macrophages with polarized phenotypes. J Biol Chem. 2012; 287: 21816-21825.
- 118.Tatano Y, Shimizu T, Tomioka H. Unique macrophages different from 36 M1/M2 macrophages inhibit T cell mitogenesis while upregulating Th17 polarization. Sci Rep. 2014; 4: 4146.
- 119.Lim SY, Yuzhalin AE, Gordon Weeks AN, et al. Tumorinfiltrating monocytes/macrophages promote tumor invasion and migration by up-regulating S100A8 and S100A9 expression in cancer cells. Oncogene. 2016; 35: 5735-5745.
- 120.Park JY, Sung JY, Lee J, et al. Polarized CD163 + tumorassociated macrophages are associated with increased angiogenesis and CXCL12 expression in gastric cancer. Clin Res Hepatol Gastroenterol. 2016; 40: 357-365.
- 121.Hu WQ, Fang M, Zhao HL, et al. Tumor invasion unit in gastric cancer revealed by QDs-based in situ molecular imaging and multispectral analysis. Biomaterials. 2014; 35: 4125-4132.
- 122.Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. Cell. 2010; 141: 39-51.
- 123.Peng C, Liu J, Yang G, et al. Lysyl oxidase activates cancer stromal cells and promotes gastric cancer progression: quantum dot-based identification of biomarkers in cancer stromal cells. Int J Nanomedicine. 2017; 13: 161-174.
- 124.Mantovani A, Sica A, Locati M. Macrophage polarization comes of age. Immunity. 2005; 23: 344-346.
- 125.Walkey CD, Olsen JB, Guo H, et al. Nanoparticle Size and Surface Chemistry Determine Serum Protein Adsorption and Macrophage Uptake. J Am Chem Soc. 2012; 134: 2139-2147.
- 126.Moreno JL, Mikhailenko I, Tondravi MM, et al. IL-4 promotes the formation of multinucleated giant cells from macrophage precursors by a STAT6-dependent homotypic mechanism contribution of E-cadherin. J Leukoc Biol. 2007; 82: 1542- 1553.
- 127.Montaner LJ, da Silva RP, Sun J, et al. Type 1 and type 2 cytokine regulation of macrophage endocytosis differential activation by IL-4/IL-13 as opposed to IFN- $\gamma$  or IL-10. J Immunol. 1999; 162: 4606-4613.
- 128.Zhang M, Hutter G, Kahn SA, et al. Anti-CD47 Treatment Stimulates Phagocytosis of Glioblastoma by M1 and M2 Polarized Macrophages and Promotes M1 Polarized Macrophages *in vivo*. PLoS ONE. 2016; 11: 0153550.
- 129.Prinz M, Priller J. Microglia and brain macrophages in the molecular age from origin to neuropsychiatric disease. Nat Rev Neuroscience. 2014; 15: 300-312.
- 130.Hu X, Li P, Guo Y, et al. Microglia/macrophage polarization dynamics reveal novel mechanism of injury expansion after focal cerebral ischemia. Stroke. 2012; 43: 3063-3070.

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